# Improved T Cell Activation Bioassays for Development of Bispecific Antibodies and Engineered T Cell Immunotherapies

Pete Stecha, Denise Garvin, Jim Hartnett, Frank Fan, Mei Cong and Zhi-jie Jey Cheng

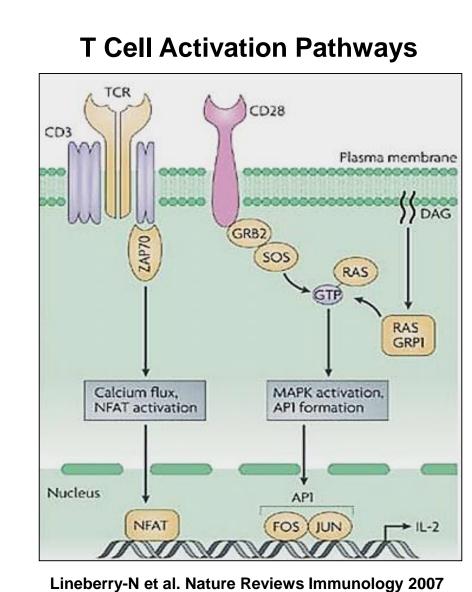
Promega Corporation, 2800 Woods Hollow Rd, Madison, WI 53711



#### 1. Introduction

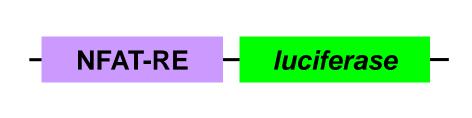
Immunotherapy aims to boost a patient's own immune system to fight disease. In recent years, a variety of immunotherapy strategies aimed at inducing, strengthening or engineering T cell responses have emerged as promising approaches for the treatment of cancer and autoimmune disease.

Here we describe a platform of T cell activation bioassays for the development of CD3 bispecific antibodies and engineered T cell immunotherapies.

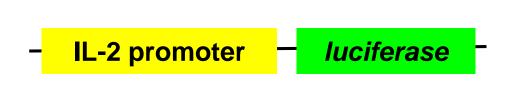


Specifically, we developed two bioluminescent reporter-based bioassays to measure T cell activation via CD3 (NFAT-RE) or CD3 + CD28 (IL-2 promoter). These bioassays include the following:

TCR/CD3 (NFAT) effector cells: Jurkat cells engineered with an NFAT-RE driving luciferase expression. Responds to CD3, but not CD28 stimulation.



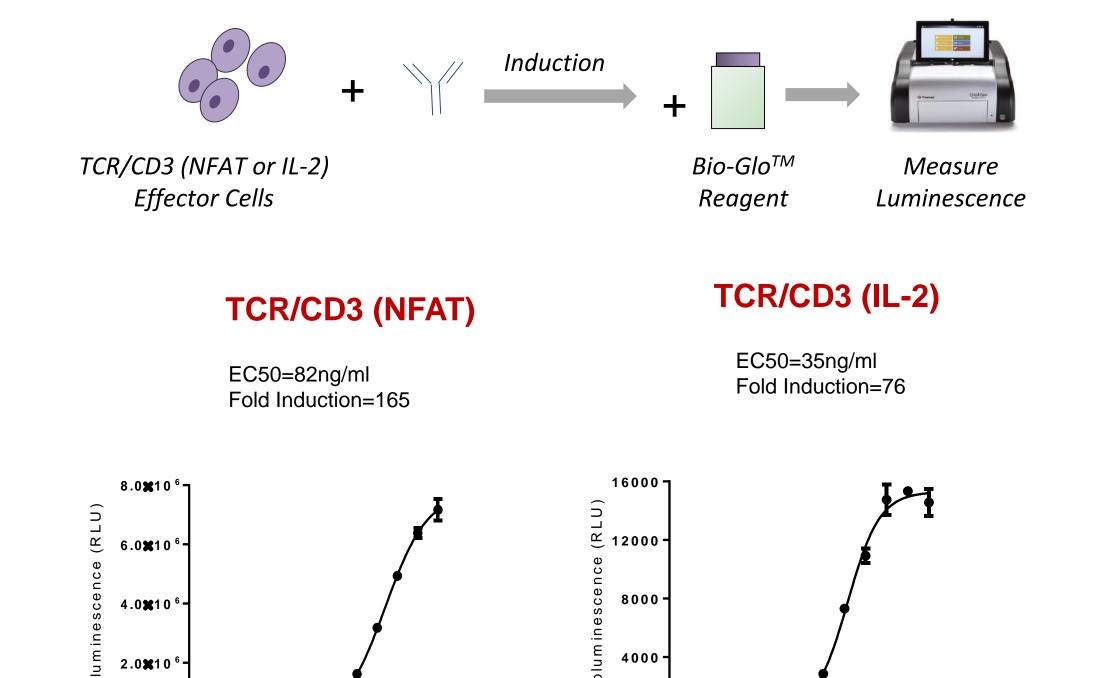
TCR/CD3 (IL-2) effector cells: Jurkat cells engineered with an IL-2 promoter driving luciferase expression. Responds to CD3 and CD3+CD28 stimulation.



Log [anti-CD3 Ab], g/m

# 2. Both TCR/CD3 (NFAT) and TCR/CD3 (IL-2) Cells Respond to TCR/CD3 Stimulation

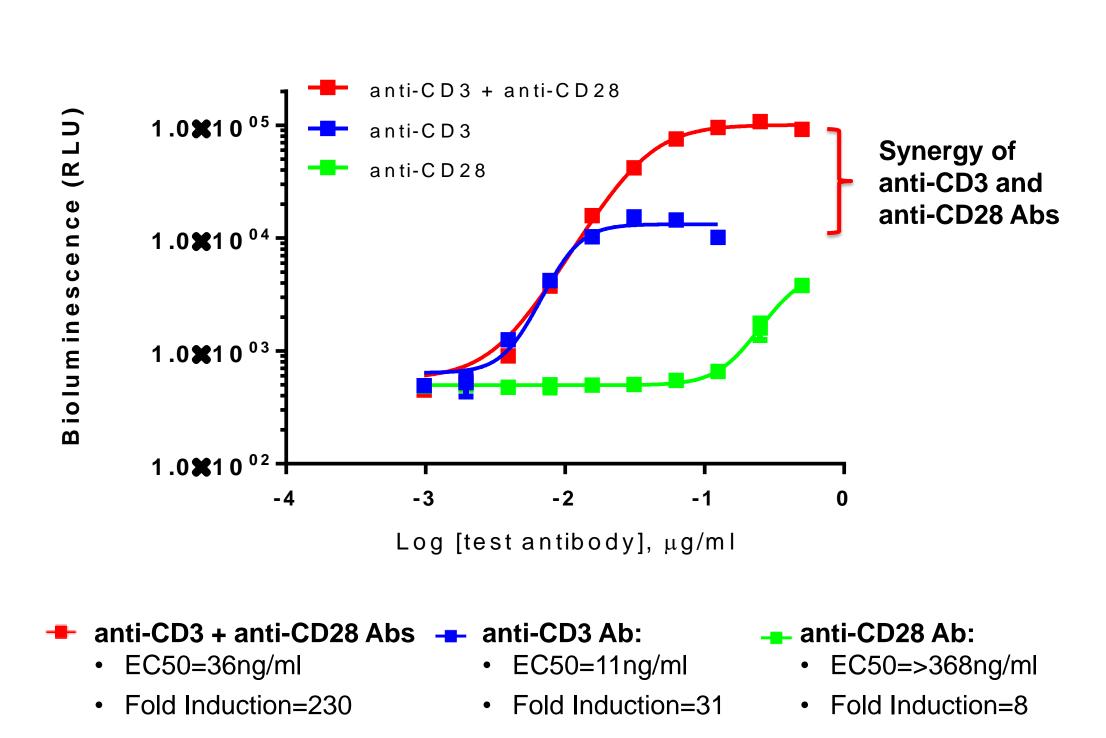
#### **T Cell Activation Protocol**



TCR/CD3 (NFAT) (Left) and TCR/CD3 (IL-2) (Right) effector cells were stimulated with increasing concentrations of an anti-CD3 Ab.

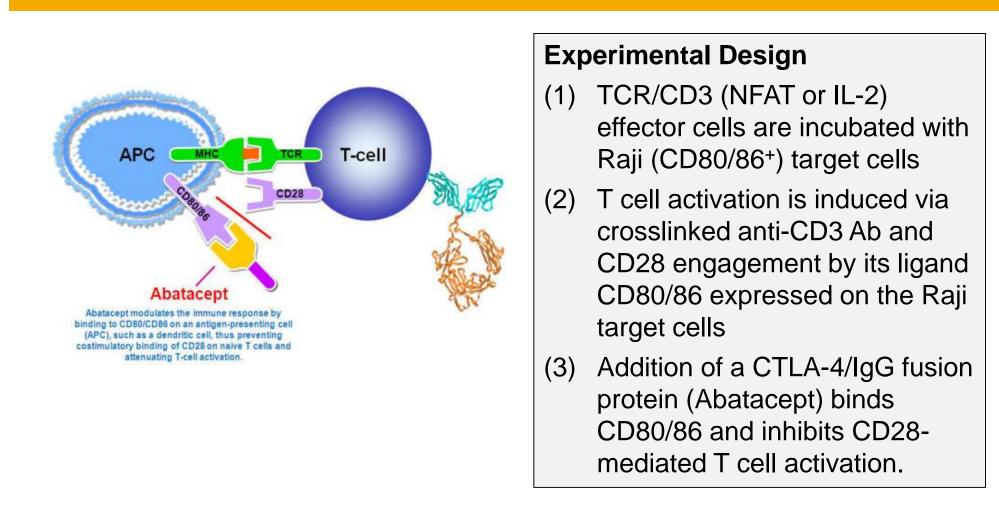
Log [anti-CD3 Ab], g/ml

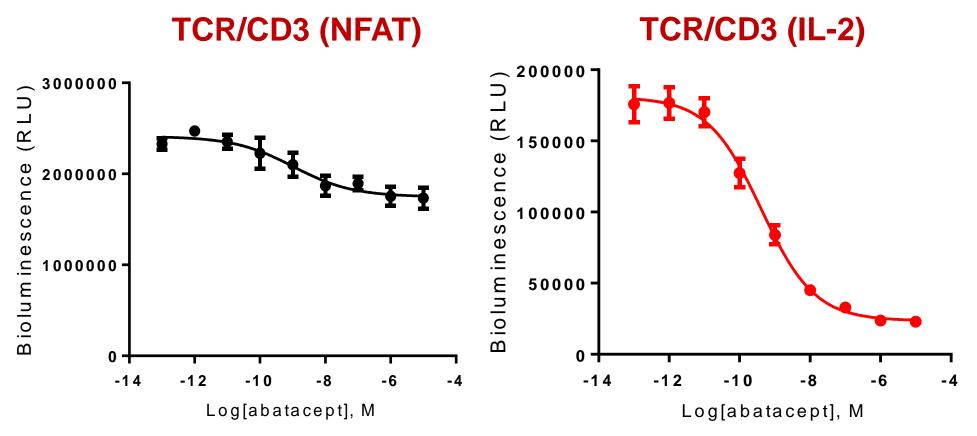
## 3. T Cell CD3+CD28 Co-stimulation Measured using TCR/CD3 (IL-2) Effector Cells



TCR/CD3 (IL-2) effector cells were stimulated with increasing concentrations of anti-CD28, anti-CD3 or a combination of anti-CD3+anti-CD28 Abs, as indicated.

#### 4. Abatacept Modulates CD3+CD28 T Cell **Activation using TCR/CD3 (IL-2) Cells**

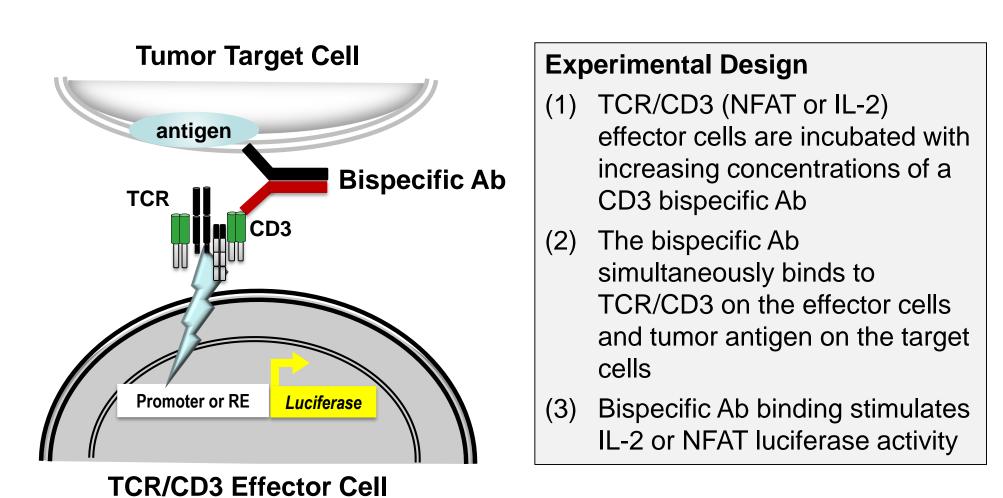




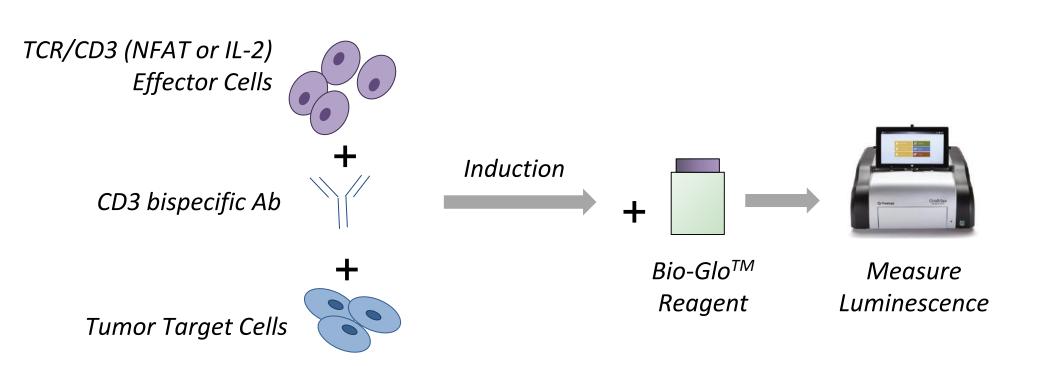
Increasing concentrations of Abatacept were added to either TCR/CD3 (NFAT) or TCR/CD3 (IL-2) effector cells, as indicated. Abatacept induced a significant decrease in TCR-mediated luciferase activity in TCR/CD3 (IL-2) effector cells compared to TCR/CD3 (NFAT) effector cells. This is expected because CD28 functions independently of the NFAT response element (see Introduction).

#### 5. Measurement of CD3 Bispecific Antibody **Activity: Assay Design and Protocol**

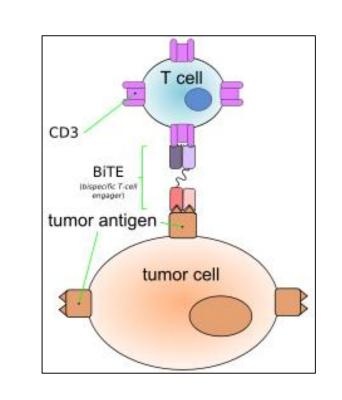
#### **Assay Design for Measuring CD3 Bispecific Antibody Activity**



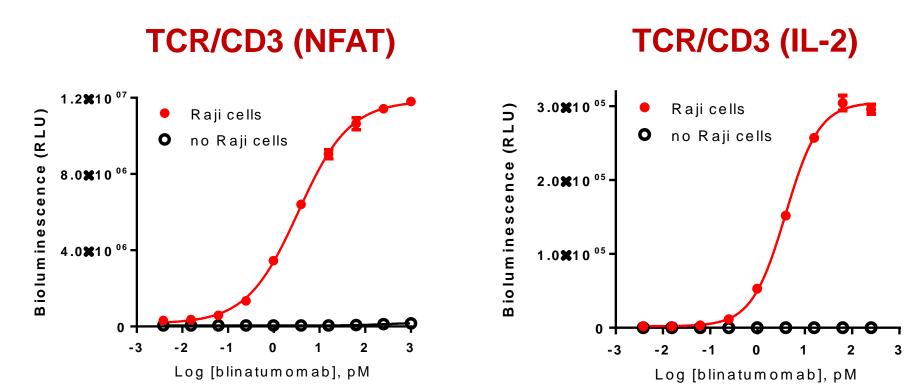
#### **Assay Protocol for Measuring CD3 Bispecific Antibody Activity**



#### 6. Analysis of Blinatumomab CD3 x CD19 **Bispecific Antibody Activity**



Blinatumomab belongs to a class of constructed monoclonal antibodies, bi-specific T-cell engagers (BiTEs), that exert action selectively and direct the human immune system to act against tumor cells. Blinatumomab specifically targets the CD19 antigen present on B cells.



Increasing concentrations of Blinatumomab were added to either TCR/CD3 (IL-2) or TCR/CD3 (NFAT) effector cells, as indicated. Blinatumomab induced a dose-dependent increase in luciferase in both TCR/CD3 (IL-2) and TCR/CD3 (NFAT) effector cells in the presence of Raji (CD19+) target cells. No response was detected in the absence of Raji (CD19+) target cells.

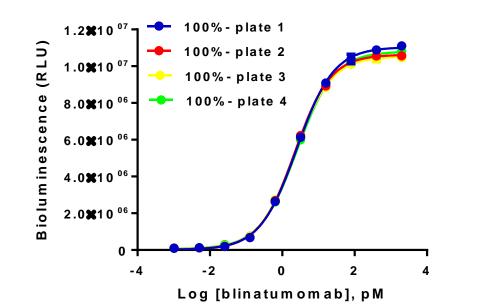
### 7. Assay Qualification with Blinatumomab: **Assay Precision, Accuracy and Linearity**

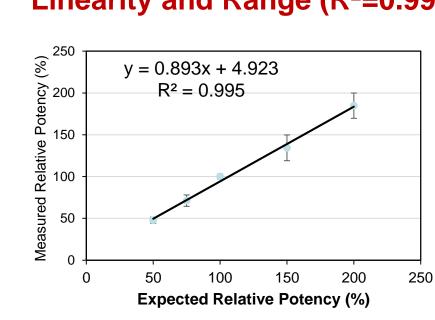
#### **Accuracy and Intermediate Precision (N = 6)**

Measured

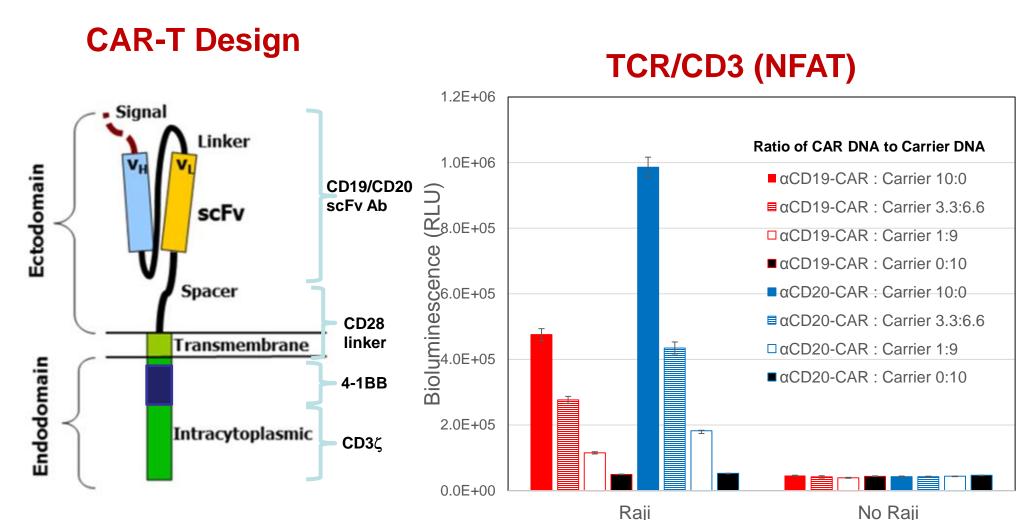
Assay Qualification Design:	Expected Relative Potency	Assay	Analyst	Measured Relative Potency %	Mean %	SD%	Accuracy Recovery %	Precisio n RSD %
Two analysts	50% 75%	1	1	45.3	71.2	3.6 6.8	95.7 95.0	7.6 9.5
		2	1	45.0				
Three days		3	1	45.5				
Four plates per day		4	2	46.2				
<ul><li>100% vs 50%</li><li>100% vs 75%</li></ul>		5	2	52.2				
		6	2	52.8				
		1	1	62.2				
<ul> <li>100% vs 150%</li> </ul>		2	1	73.9				
<ul> <li>100% vs 200%</li> <li>Data shown are generated using</li> </ul>		3	1	63.3				
		4	2	78.4				
		5	2 2	75.9 73.6				
	150%	1	1	144.5	134.4	15.5	89.6	11.5
		2	1 1	143.5				
9		3	1 1	121.4				
TCR/CD3 (NFAT)		4	2	108.8				
effector cells,		5	2	143.4				
binatumomab,		6	2	144.7				
and Raji (CD19+) target cells.	200%	1	1	174.9	184.8	15.0	92.4	8.1
		2	1	195.5				
		3	1	179.8				
		4	2	162.0				
		5	2	198.9				
		6	2	198.0				
						Overall	93.2	9.2

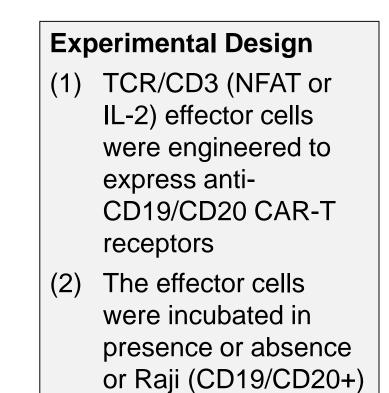
#### **Linearity and Range (R<sup>2</sup>=0.995)** Repeatability (%CV) = 3.01%



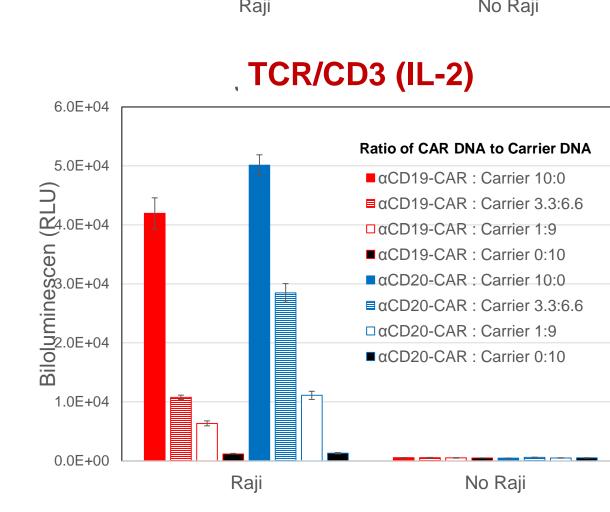


## 8. Measurement of Chimeric Antigen Receptor T (CAR-T) Cell Activity





target cells



Serial dilutions of anti-CD19/CD20 CAR-T effector cells were incubated in the presence and absence of Raji (CD19/CD20+) target cells. Luciferase activity was detected in the presence of Raji cells, but not with the CAR-T effector cells alone.

#### 9. Conclusions

We have developed a platform of T cell activation bioassays that incorporate a bioluminescent reporter-based readout of T cell activation via CD3 (NFAT-RE) or CD3 + CD28 (IL-2 promoter). These assays reflect the mechanisms of action of biologics designed to engage, recruit, and stimulate T cell activation to attack target disease cells. Specific applications include measurement of anti-CD3 bispecific Ab and CAR-T cell activity.

The bioassays provide the following:

# Mechanism of action (MOA)-based measure of biologics activity

- Specific measure of CD3 or CD3 + CD28 T cell activation pathways
- Quantitative measure of anti-CD3 Ab and bispecific Ab potency

# Consistent and reliable measure of biologics activity

- Demonstrated precision, accuracy, reproducibility, robustness
- All assays can be used as "Thaw-and-use" cell format, no cell culture required
- Functional performance suitable for development into potency, stability, and NAb assays (data not shown)

#### **Easy-to-implement**

- Rapid and convenient workflow
- Amenable to standard 96-well and 384-well plate formats